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SESSILE ALGAE

IN LOWER SECTIONS OF THE MAIN TRIBUTARIES OF THE WIDAWKA RIVER*

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ABSTRACT: Sessile algae were collected from lower sections of the rivers: Rakówka, Pilsia, Krasawa, Grabia and Nieciecz. The investigated rivers lie within a cone of depression formed due to the construction of the lignite open mine Bełchatów. Quantitative and qualitative composition of algal communities was determined in the samples. In total, 417 taxa of algae and 3 taxa of bacteria were recorded - among them: 153 taxa from the Rakówka River, 211 taxa from the Pilsia River, 222 taxa from the Krasawa River, 220 taxa from the Grabia River, and 234 taxa from the Nieciecz River.

Contents

1. Introduction
2. Description of study area
3. Material and methods
4. Results and discussion
 - 4.1. Qualitative composition
 - 4.2. Quantitative composition

* The studies were carried out within the project WE 10.2.05 "Protection of the Environment in Large Industrial Agglomerations".

5. Conclusions
6. References
7. Streszczenie

1. INTRODUCTION

Algae of the water-courses of the area of Bełchatów Industrial Region were already investigated in the main river of the region - Widawka River, in its tributary - Rakówka River and in the drainage channels of the lignite mine (K a d ł u b o w s k a et al. 1981). P a w ł o w s k i's work (1969) contains data about algae occurring in the Grabia River syrtion. A list of algological studies on other water reservoirs in this area can be found in the paper by C z y ż e w s k a and O l a c z e k (1983).

Within the framework of the research project WE 10.2.05 there were undertaken investigations on sessile algae in the Widawka River and its main tributaries i.e. the Rakówka River, the Pilsia River, the Krasawa River, the Grabia River, the Nieciecz River and in the channels draining the lignite mine area. The first study on the above subject concerned sessile algae in the Widawka River (L i g o w s k i in press b).

The aim of the present study is to determine qualitative and quantitative composition of sessile algal communities in lower sections of the five above mentioned main tributaries of the Widawka River.

2. DESCRIPTION OF STUDY AREA

The investigated rivers are situated in the subprovince of Central Poland Lowland belonging to the province of Central European Lowland. Springs and upper sections of all investigated tributaries of the Widawka River are situated on Bełchatów Upland, and their middle and lower sections flow across Szczerców Valley. An exception here is the Grabia River, whose middle section flows across the Łask Upland (K o n d r a c k i 1978).

Almost the entire Widawka River along with its tributaries:

Rakówka, Pilsia, Krasawa and Nieciecz lie within a cone of depression formed due to the construction of the lignite open mine - Bełchatów (Czyżewska and Olaczek 1983).

Among the investigated tributaries of the Widawka River, the Rakówka River is a most polluted one (Hereźniak 1972), the Pilsia River is somewhat less polluted, especially in its lower section. Waters of such rivers as Grabia, Krasawa and Nieciecz are relatively clean (Szarkowski 1977, Maksymiuk 1979).

The Rakówka River is a right side tributary of the Widawka River (Fig. 1). The Rakówka River receives sewage from the towns of Bełchatów and Wola Krzysztoporska (Szarkowski 1977, Maksymiuk 1979). Its length is 20.4 km, surface of the drainage basin amounts to 147.9 km^2 and mean gradient is 1.79%. (Maksymiuk 1980). At the place where samples were collected, near the bridge on the road from Słok to Bełchatów, the river is 4-5 m wide, and up to 0.5 m deep, its bottom is covered with silt and stones.

The Pilsia River is a right side tributary of the Widawka River (Fig. 1). In its spring section, the sewage from Żelów causes a considerable pollution of the water, while in its lower section the degree of pollution is smaller (Maksymiuk 1979). The Pilsia River is 30.0 km long, and its mean gradient is 1.95%. Its drainage basin amounts to 192.3 km^2 (Maksymiuk 1980). At the station where samples were collected, i.e. near the road from Szczerców to Widawa, the river is about 4 m wide and its bottom is covered with sand and silt.

The Krasawa (Krasówka) River is a left side tributary of the Widawka River (Fig. 1). Its length is 34.2 km, mean gradient - 1.94%, and its drainage basin - 196 km^2 (Maksymiuk 1980). The samples were collected near the road from Szczerców to Wieruln, where the river is ca. 5 m wide and its bottom is covered with sand and stones.

The Grabia River is the largest tributary of the Widawka River (Fig. 1). Its length is 82.1 km, its mean gradient amounts to 1.07%, and its drainage basin amounts to 819.5 km^2 (Maksymiuk 1980). The samples were collected near the road from Górki Grabieńskie to Siedlce. At this station the river is ca.

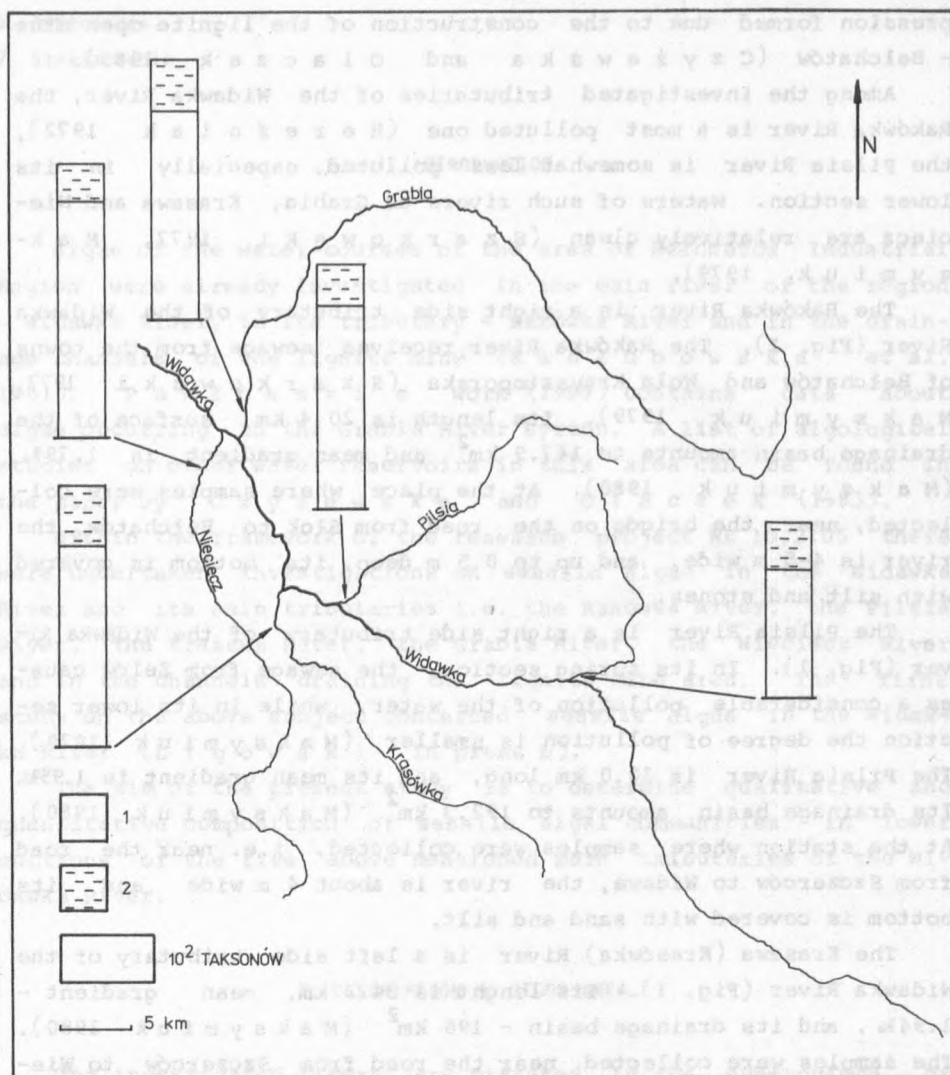


Fig. 1. Number of taxa of the sessile algae in lower sections of the main tributaries of the Widawka River

20 m wide and has a bottom covered with sand and stones and with silt in calm places.

The Nieciecz River - a left side tributary of the Widawka River (Fig. 1) empties into Widawka near the town of Widawa, where

the samples were taken. Its length is 49.5 km, mean gradient - 1.95%, and drainage basin - 261.5 km² (M a k s y m i u k 1980). At the station where the samples were collected the river is about 15 m wide, and its bottom is covered with sand and stones.

The total drainage basin of the rivers: Rakówka, Pilsia, Krasawa, Grabia and Nieciecz amounts to 1617 km², which accounts for 66.6% of the total drainage basin of the Widawka River.

3. MATERIAL AND METHODS

The samples of permanently or accidentally sessile algae were collected near the bank of lower sections of the rivers: Rakówka, Pilsia, Krasawa, Grabia and Nieciecz on 15th May, 16th June, 15th July and 19th November 1982. Algae from stones and macrophytes were scraped off, and silt from the river's bottom was collected directly into submerged bottles. Immediately after their collecting, the samples were fixed by 2% formaline. A part of the samples were used to make permanent diatom slides in pleurax (S i e m i ń s k a 1964). In order to identify taxa, all specimens of diatoms present in permanent slides were checked. During the species determination procedure over 300 successive diatom valves were counted. These data were helpful in calculating percentage composition of diatom communities (C h o l n o k y 1968, K a l b e 1973). Algae belonging to other systematic groups were determined in water slides with glycerine added (S t a r m a c h 1955). The quantitative composition of algal community was established in the following way: the slide was done from a sample diluted in 1:4 ratio; the slide contained 0.05 cm³ of this diluted sample and counting was performed in one-eighth part of a slide. The numbers specimens of particular diatom species were obtained taking into account percentage composition of diatom community determined on the basis of permanent slides. The algae whose number of cells accounted for over 5% the overall number of cells were recognized to be dominant.

While identifying taxa, the following papers were mainly utilized: B a r t a et al. (1976), C l e v e - E u l e r

Table 1 (contd.)

1	2	3	4	5	6
(Kützing) Bornet et Flahault				+	
<i>Cylindrospermum</i> sp.			+		
<i>Gloeocapsa turgida</i> (Kützing) Hollerbach		+	+	+	
<i>Gomphosphaeria compacta</i> (Lemmermann) Strom			+	+	+
<i>G. lacustris</i> Chodat			+		
<i>Lyngbya cryptovaginata</i> Schkorbatoff			+	+	+
<i>L. hieronymusii</i> Lemmermann			+	+	+
<i>L. limetica</i> Lemmermann	++		+		+
<i>Merismopedia glauca</i> (Ehrenberg) Nägeli				++	+
<i>M. tenuissima</i> Lemmermann			++		+
<i>Microcystis aeruginosa</i> Kütz- ing		++			
<i>M. incerta</i> (Lemmermann) Star- mach		+			
<i>Oscillatoria agardhii</i> Gomont			+++	+	+
<i>O. amoena</i> (Kützing) Gomont			+		
<i>O. brevis</i> (Kützing) Gomont	+				
<i>O. chalybaea</i> (Mertens) Gomont	+				
<i>O. limosa</i> Agardh	++	+	+++	+	++++
<i>O. pseudogeminata</i> G. Schmid			+++		++
<i>O. sancta</i> (Kützing) Gomont	+		+		
<i>O. tenuis</i> Agardh	++		++		+
<i>O. terebriformis</i> Agardh				+	
<i>Oscillatoria</i> sp.	+	+			
<i>Pseudanabaena schmidlei</i> Jaag			++		
<i>Pseudanabaena</i> sp.	+				
<i>Schizothrix</i> sp.	+				
<i>Spirulina maior</i> Kützing				+	
<i>Spirulina tenuissima</i> Kützing			+		

Table 1 (contd.)

1	2	3	4	5	6
<i>Pyrrophyta</i>					
<i>Gymnodinium</i> sp.			++		
<i>Peridinium</i> sp.			+		
<i>Euglenophyta</i>					
<i>Euglena spirogyra</i> Ehrenberg	+				+
<i>Euglena</i> sp.	++++	+	+	++	+++
<i>Lepocinclis ovum</i> (Ehrenberg)					
Minkiewicz				+	+
<i>Phacus acuminatus</i> Stokes	+	++	++		+
<i>Ph. acuminatus</i> var. <i>megaparamylica</i> (Roll) Huber-Post.					+
<i>Ph. caudatus</i> Hübner					++
<i>Ph. caudatus</i> var. <i>tenuis</i> Swirenko	+				
<i>Ph. longicauda</i> (Ehrenberg) Dujardin		+			
<i>Ph. mirabilis</i> Pochman		+			
<i>Ph. orbicularis</i> Hübner		+	+		+
<i>Ph. pleuronectes</i> (Ehrenberg) Dujardin	++	+			
<i>Ph. pyrum</i> Ehrenberg				+	
<i>Trachelomonas hispida</i> (Perty) Stein	+		++	+	++
<i>T. hispida</i> var. <i>coronata</i> Lemmermann		+	+	+	+
<i>T. oblonga</i> Lemmermann	+		++	+	
<i>T. volvocina</i> Ehrenberg	++		+	++	++
<i>Chrysophyceae</i>					
<i>Dinobryon sertularia</i> var. <i>protuberans</i> (Lemm.) Krieger		+			
<i>Mallomonas</i> sp.	+	+			
<i>Bacillariophyceae</i>					
<i>Achnanthes affinis</i> Grun.		+	++	+	++
<i>A. clevei</i> Grun.					+

Table 1 (contd.)

1	2	3	4	5	6
<i>A. clevei</i> var. <i>rostrata</i> Hust.					+
<i>A. delicatula</i> (Kütz.) Grun.		+	++	++	++++
<i>A. exilis</i> Kütz.			+		
<i>A. exigua</i> Grun.					+
<i>A. exigua</i> var. <i>heterovalvata</i> Krasske		+			
<i>A. hungarica</i> Grun.	++	++++	+	+	+
<i>A. lanceolata</i> (Bréb.) Grun.	+++	++++	+	++++	+++
<i>A. lanceolata</i> f. <i>capitata</i> O. Müll.			+	+	++
<i>A. lanceolata</i> var. <i>elliptica</i> Cl.	++++		++	+++	++++
<i>A. lanceolata</i> var. <i>rostrata</i> (Østr.) Hust.	+	+	++++	++	++
<i>A. lanceolata</i> f. <i>ventricosa</i> Hust.	++	+			+
<i>A. linearis</i> (W. Sm.) Grun.		+	+	+	++
<i>A. microcephala</i> (Kütz.) Grun.				+	+
<i>A. minutissima</i> Kütz.				+	
<i>A. peragallii</i> Brun et Héríb.		++	+		
<i>A. saxonica</i> Krasske					+
<i>Amphipleura pellucida</i> Kütz.			+		
<i>Amphora normanii</i> Rabh.					+
<i>A. ovalis</i> Kütz.	+++	++++	++	++++	++++
<i>A. ovalis</i> f. <i>gracilis</i> (Ehr.) Cl.					+
<i>A. ovalis</i> var. <i>libyca</i> (Ehr.) Cl.			+	++	
<i>A. ovalis</i> var. <i>pediculus</i> Kütz.		+	++++	++	++
<i>A. veneta</i> Kütz.			+		+
<i>Anomoeoneis sphaerophora</i> (Kütz.) Pfitz.				+	++
<i>Asterionella formosa</i> Hass.		+	+++	+	+
<i>Caloneis amphisbaena</i> (Bory) Cl.	+++	+	+++	+++	+++

Table 1 (contd.)

1	2	3	4	5	6
<i>C. bacillum</i> (Grun.) Mer.		++	++++	+	++
<i>C. bacillum</i> var. <i>lancettula</i> (Schulz) Hust.		+	++	+	+++
<i>C. silicula</i> (Ehr.) Cl.		+ +	++	+++	+
<i>C. silicula</i> var. <i>gibberula</i> (Kütz.) Grun.	+		+	+	
<i>C. silicula</i> var. <i>truncatula</i> Grun.		++		+	+
<i>C. silicula</i> var. <i>ventricosa</i> (Ehr.) Donk.				+	+
<i>Compylodiscus noricus</i> var. <i>hibernica</i> (Ehr.) Grun.		+	+	+	+
<i>Cocconeis diminuta</i> Pant.	+		+	+	++++
<i>C. disculus</i> (Schum.) Cl.		++	+		
<i>C. pediculus</i> Ehr.			++++	+++	++++
<i>C. placentula</i> Ehr.	++++	++++	++++	+++	++++
<i>C. placentula</i> var. <i>euglypta</i> (Ehr.) Cl.		++++	++++	+	++++
<i>C. placentula</i> var. <i>intermedia</i> (Héríb. et Perag.) Cl.				+	
<i>C. placentula</i> var. <i>lineata</i> (Ehr.) Cl.				++	+
<i>C. thumensis</i> Mayer		+	+		+
<i>Cyclotella comta</i> (Ehr.) Kütz.	+		+	+	+
<i>C. kützingiana</i> Thw.			+	+	+
<i>C. meneghiniana</i> Kütz.	++++	++++	++++	+++	+++
<i>Cymatopleura elliptica</i> (Bréb.) W. Sm.		++	++	+	++
<i>C. elliptica</i> var. <i>hibernica</i> (W. Sm.) Hust.				+	
<i>C. elliptica</i> var. <i>nobilis</i> (Hantzsch) Hust.				+	+
<i>C. solea</i> (Bréb.) W. Sm.	++++	++++	+++	+++	++++
<i>C. solea</i> var. <i>apiculata</i> (W. Sm.) Ralfs		+++	++	++	+++

Table 1 (contd.)

1	2	3	4	5	6
<i>C. solea</i> var. <i>gracilis</i> Grun.		+		+	+
<i>Cymbella affinis</i> Kütz.			+		+
<i>C. amphicephala</i> Näg.			+		
<i>C. aspera</i> (Ehr.) Cl.	+	+	+	+	+
<i>C. cistula</i> (Hemp.) Grun.		+	+	+	+
<i>C. cistula</i> var. <i>maculata</i> (Kütz.) VH			+		
<i>C. cuspidata</i> Kütz.	+				
<i>C. ehrenbergii</i> Kütz.		+	+	+	+
<i>C. lanceolata</i> (Ehr.) VH		+	+	+	+
<i>C. naviculiformis</i> Auersw.	+	+	+	+	+
<i>G. parva</i> (W. Sm.) Cl.		+			
<i>C. prostrata</i> (Berkeley) Cl.				+	
<i>C. reinhardtii</i> Grun.					+
<i>C. sinuata</i> Greg.	+	+	+	+	+
<i>C. tumida</i> (Bréb.) VH			+		
<i>C. turgida</i> (Greg.) Cl.	+				
<i>C. ventricosa</i> Kütz.	+	+	+	+	+
<i>Diatoma elongatum</i> (Lyngb.) Ag.			+	+	
<i>D. vulgare</i> Bory	+	+	+	+	+
<i>Diploneis oculata</i> (Bréb.) Cl.			+		
<i>D. ovalis</i> (Hilse) Cl.	+	+	+		+
<i>D. smithii</i> (Bréb.) Cl.	+	+			
<i>D. smithii</i> var. <i>pumila</i> (Grun.) Hust.				+	
<i>Epithemia sorex</i> Kütz.	+	+		+	+
<i>E. turgida</i> (Ehr.) Kütz.	+	+	+	+	+
<i>E. turgida</i> var. <i>granulata</i> (Ehr.) Grun.		+	+	+	
<i>E. zebra</i> (Ehr.) Kütz.	+	+	+	+	+
<i>E. zebra</i> var. <i>porcellus</i> (Kütz.) Grun.	+	+	+		
<i>E. zebra</i> var. <i>saxonica</i> (Kütz.) Grun.		+	+		+

Table 1 (contd.)

1	2	3	4	5	6
<i>Eunotia arcus</i> Ehr.	+	+			+
<i>E. arcus</i> var. <i>fallax</i> Hust.			+		
<i>E. diodon</i> Ehr.		+			
<i>E. lunaris</i> (Ehr.) Grun.	++++	++	+	++	++
<i>E. lunaris</i> var. <i>capitata</i> Grun.		+		+	
<i>E. monodon</i> var. <i>maior</i> (W. Sm.) Hust.		+			
<i>E. pectinalis</i> var. <i>minor</i> (Kütz.) Rabh.				+	
<i>E. praerupta</i> Ehr.		++	++		
<i>Fragilaria bicapitata</i> Mayer		++	+		
<i>F. brevistriata</i> Grun.	+	+++	+++	+	++
<i>F. capucina</i> Desm.	+				
<i>F. capucina</i> var. <i>lanceolata</i> Grun.			+		
<i>F. capucina</i> var. <i>mesolepta</i> Rabh.		+	+		
<i>F. construens</i> (Ehr.) Grun.	++++	++++	++++	+	++
<i>F. construens</i> var. <i>binodis</i> (Ehr.) Grun.	++++	++++	+++	++	++
<i>F. construens</i> var. <i>subsalina</i> Hust.	+	+			
<i>F. construens</i> var. <i>venter</i> (Ehr.) Grun.	++	++++	++++	+	++++
<i>F. crotonensis</i> Kitt.			+		+
<i>F. inflata</i> (Heid.) Hust.			+		
<i>F. intermedia</i> Grun.	++	++++	++++	+++	+
<i>F. leptostauron</i> (Ehr.) Hust.		++	++	+	+
<i>F. leptostauron</i> var. <i>dubia</i> Grun.			+		
<i>F. pinnata</i> Ehr.	+	+	++		
<i>F. pinnata</i> var. <i>lancettula</i> (Schum.) Hust.				+	

Table 1 (contd.)

1	2	3	4	5	6
<i>F. vaucheriae</i> (Kütz.) Boye Pet.	+			+	
<i>F. virescens</i> Ralfs.				+	
<i>F. virescens</i> var. <i>capitata</i> Østr.	+	+			
<i>F. virescens</i> var. <i>elliptica</i> Hust.	+			+	
<i>F. virescens</i> var. <i>oblongella</i> Grun.				+	
<i>Frustulia vulgaris</i> (Thw.) De Toni	++	+++	++++	+++	++++
<i>Gomphonema acuminatum</i> Ahr.	+	+++	+		+++
<i>G. acuminatum</i> var. <i>brebissonii</i> (Kütz.) Cl.	+	+	+		
<i>G. acuminatum</i> var. <i>coronatum</i> (Ehr.) W. Sm.		++	+	++	+++
<i>G. acuminatum</i> var. <i>trigonocephalum</i> (Ehr.) Grun.	+	++			+
<i>G. angustatum</i> (Kütz.) Rabh.	+++	++++	++	++	++
<i>G. angustatum</i> var. <i>productum</i> Grun.				+	++
<i>G. angustatum</i> var. <i>sacrophagus</i> (Greg.) Grun.			+	++	+
<i>G. angustatum</i> var. <i>undulatum</i> Grun.					+
<i>G. augur</i> Ehr.		+			
<i>G. augur</i> var. <i>gautieri</i> V.H.					+
<i>G. constrictum</i> Ehr.	++	++	++	+++	+++
<i>G. gracile</i> Ehr.			++		+
<i>G. intricatum</i> Kütz.		+	+	+	+
<i>G. intricatum</i> var. <i>pumilum</i> Grun.					+
<i>G. intricatum</i> var. <i>vibrio</i> (Ehr.) Cl.		+			
<i>G. lanceolatum</i> Ehr.	+	++			+

Table 1 (contd.)

1	2	3	4	5	6
<i>G. longiceps</i> var. <i>subclavatum</i> Grun.		+	+		
<i>G. olivaceum</i> (Lyngb.) Kütz.			++++	++	++++
<i>G. olivaceum</i> var. <i>calcareum</i> Cl.			+	+	+
<i>G. olivaceum</i> var. <i>minutissimum</i> Hust.			+		
<i>G. parvulum</i> (Kütz.) Grun.	++++	++++	+++	+++	++++
<i>G. parvulum</i> var. <i>micropus</i> (Kütz.) Cl.			+	++	+
<i>G. parvulum</i> var. <i>subellipticum</i> Cl.			+	+	
<i>Gyrosigma acuminatum</i> (Kütz.) Rabh.					++
<i>G. attenuatum</i> (Kütz.) Rabh.	+	++	++	+	++
<i>G. kützingii</i> (Grun.) Cl.		++	+	+	+++
<i>Hantzschia amphioxys</i> f. <i>capitata</i> Hust.	+	+++			++
<i>Melosira arenaria</i> Moore				+	
<i>M. granulata</i> (Ehr.) Ralfs		++			+
<i>M. granulata</i> var. <i>angustissima</i> (O. Müll.) Hust.		+	+		+
<i>M. islandica</i> O. Müll.		+			
<i>M. italica</i> (Ehr.) Kütz.	+	+	+	+	+
<i>M. italica</i> f. <i>curvata</i> (Pant.) Hust.					+
<i>M. italica</i> var. <i>tenuissima</i> (Grun.) O. Müll.		+			
<i>M. varians</i> Ag.	++	+++	++++	+	++++
<i>Meridion circulare</i> Ag.	++++	++++	++++	+++	++++
<i>M. circulare</i> var. <i>constricta</i> (Ralfs) V.H.	+	+		+	+
<i>Navicula amphibola</i> Cl.	+	++			
<i>N. anglica</i> Ralfs	+		+		+
<i>N. bacilliformis</i> Grun.	+	++	+		+

Table 1 (contd.)

1	2	3	4	5	6
<i>N. bacillum</i> Ehr.	+	+	++	++++	++ ++
<i>N. bacillum</i> var. <i>gregoryana</i> Grun.					
<i>N. bacillum</i> var. <i>rostrata</i> Mayer					
<i>N. binodis</i> Ehr.		+			
<i>N. cari</i> Ehr.		++	+++	+	+
<i>N. cincta</i> (Ehr.) Kütz.	+	++++		+	++++
<i>N. cincta</i> var. <i>heufleri</i> Grun.					+
<i>N. clementis</i> Grun.			++	+	+
<i>N. cocconeiformis</i> Greg.					+
<i>N. contenta</i> Grun.					+
<i>N. costulata</i> Grun.		+	+	+++	++ ++
<i>N. cryptocephala</i> Kütz.	+++	+++	++++	++++	++++
<i>N. cryptocephala</i> var. <i>intermedia</i> Grun.		+++	++++	+++	++++
<i>N. cryptocephala</i> var. <i>veneta</i> (Kütz.) Grun.		+	++++	++	++
<i>N. cuspidata</i> Kütz.	+	+	+	+++	+++
<i>N. cuspidata</i> var. <i>ambigua</i> (Ehr.) Cl.	++	+	+	+	++++
<i>N. dicephala</i> (Ehr.) W. Sm.	+	+	++	++	++
<i>N. dicephala</i> var. <i>neglecta</i> (Krasske) Hust.	+	+			
<i>N. digitoradiata</i> (Greg.) A. Sm.		+		+	++
<i>N. exigua</i> (Greg.) O. Müll.	++	+++	++++	++++	++++
<i>N. gastrum</i> Ehr.	+	+++	++	++	++
<i>N. gracilis</i> Ehr.	+		++++	+++	++++
<i>N. graciloides</i> Mayer			+	+	
<i>N. gregaria</i> Donk.	+++		+++	+++	
<i>N. halophila</i> (Grun.) Cl.	++	+			
<i>N. hungarica</i> Grun.					+
<i>N. hungarica</i> var. <i>capitata</i> (Ehr.) Cl.	++++	++++	++++	++++	++++

Table 1 (contd.)

1	2	3	4	5	6
<i>N. hungarica</i> var. <i>lüneburgen-</i> <i>sis</i> Grun.				+	+
<i>N. integra</i> (W. Sm.) Ralfs	+	+	+	+	+
<i>N. lanceolata</i> (Ag.) Kütz.		+			+
<i>N. menisculus</i> Schum.		+	+	+	+
<i>N. menisculus</i> var. <i>meniscus</i> (Schum.) Hust.			+		
<i>N. minima</i> Grun.				+	
<i>N. mutica</i> Kütz.		+			+
<i>N. mutica</i> var. <i>cohnii</i> (Hilse) Grun.			+		
<i>N. mutica</i> var. <i>nivalis</i> (Ehr.) Hust.	+				
<i>N. oblonga</i> Kütz.		+	+	+	+
<i>N. oppugnata</i> Hust.		+	+	+	+
<i>N. pelliculosa</i> (Bréb.) Kütz.				+	
<i>N. placentula</i> (Ehr.) Grun.	+	+	+	+	+
<i>N. placentula</i> var. <i>jenisseje-</i> <i>sis</i> (Grun.) Meist.					+
<i>N. placentula</i> f. <i>latiuscula</i> (Grun.) Meist.				+	
<i>N. placentula</i> f. <i>rostrata</i> Mayer		+	+	+	+
<i>N. protracta</i> Grun.	+	+	+	+	+
<i>N. protracta</i> f. <i>elliptica</i> Gallik			+		
<i>N. protracta</i> f. <i>subcapitata</i> (Wisł. et Por.) Hust.				+	+
<i>N. pupula</i> Kütz.	+	+	+	+	+
<i>N. pupula</i> var. <i>capitata</i> Hust.			+	+	+
<i>N. pupula</i> var. <i>elliptica</i> Hust.					+
<i>N. pupula</i> var. <i>rectangularis</i> (Greg.) Grun.		+	+	+	+
<i>N. pygmaea</i> Kütz.			+	+	+

Table I (contd.)

1	2	3	4	5	6
<i>N. radiosa</i> Kütz.		+++	++++	+	++
<i>N. radiosa</i> var. <i>tenella</i> (Bréb.) Grun.		+	+		+
<i>N. reinhardtii</i> Grun.	++	++	++++	++++	++++
<i>N. reinhardtii</i> var. <i>gracilior</i> Grun.			+	++	+
<i>N. rhynchocephala</i> Kütz.		++++	++++	+++	++
<i>N. rotaena</i> Grun.	++ +	++++	+	++	+ +
<i>N. subhamulata</i> Grun.			++ +		+
<i>N. tuscula</i> (Ehr.) Grun.	+		+		
<i>N. viridula</i> Kütz.	+++	++++	++++	+++	++++
<i>N. viridula</i> var. <i>avenacea</i> (Bréb.) Grun.			+	+++	+++
<i>N. viridula</i> var. <i>capitata</i> Mayer		+	+	++	+
<i>N. viridula</i> var. <i>slesvicensis</i> (Grun.) Cl.	+	++ +	+	++	++
<i>Neidium affine</i> (Ehr.) Cl.					+
<i>N. affine</i> var. <i>amphirhynchus</i> (Ehr.) Cl.	++++	++	++	++++	+ +
<i>N. bisulcatum</i> (Lagerst.) Cl.				+	
<i>N. dubium</i> (Ehr.) Cl.	+	+	+++	++	++
<i>N. dubium</i> f. <i>constrictum</i> Hust.				+	+
<i>N. iridis</i> (Ehr.) Cl.					+
<i>N. iridis</i> var. <i>amphigomphus</i> (Ehr.) V.H.		+		+	
<i>N. iridis</i> var. <i>ampliatum</i> (Ehr.) Cl.		+	++	+	+
<i>N. iridis</i> f. <i>vernale</i> Reich.	+	+	++ +		++
<i>N. opulentum</i> Hust.				+	+
<i>N. productum</i> (W. Sm.) Cl.	+				++
<i>Nitzschia acicularis</i> W. Sm.	++++	+	++++	++++	++++
<i>N. acuta</i> Hantzsch			+	+	+
<i>N. amphibia</i> Grun.	+	+	+	++	++

Table 1 (contd.)

1	2	3	4	5	6
<i>N. angustata</i> (W. Sm.) Grun.					+
<i>N. apiculata</i> (Greg.) Grun.					++
<i>N. capitellata</i> Hust.	+				
<i>N. dissipata</i> (Kütz.) Grun.	+		++++	+	+
<i>N. fonticola</i> Grun.		+	+++	+	+++
<i>N. frustulum</i> (Kütz.) Grun.	++	+	++		++
<i>N. gracilis</i> Hantzsch				+	
<i>N. hantzschi</i> Rabh.					
<i>N. heidenii</i> var. <i>pamirensis</i> Boye P.		+			+
<i>N. hungarica</i> Grun.					+
<i>N. kützingiana</i> Hilse					+
<i>N. linearis</i> W. Sm.	++++	++	+	++	+
<i>N. palea</i> (Kütz.) W. Sm.	++++	++	+++	++++	++++
<i>N. palea</i> var. <i>tenuirostris</i> Grun.	+				
<i>N. parvula</i> Lewis			+		
<i>N. recta</i> Hantzsch	+		+	+	+
<i>N. sigmoidea</i> (Ehr.) W. Sm.		+	+	++	++
<i>N. sinuata</i> var. <i>tabellaria</i> Grun.					+
<i>N. stagnorum</i> Rabh.	++			+++	
<i>N. sublinearis</i> Hust.			+		
<i>N. thermalis</i> Kütz.	+		+		+
<i>N. thermalis</i> var. <i>minor</i> Hilse					+
<i>N. trybionella</i> Hantzsch					+
<i>N. trybionella</i> var. <i>debilis</i> (Arn.) Mayer					+
<i>N. vermicularis</i> (Kütz.) Grun.				++	+
<i>Opephora martyi</i> Héríb.	++++	++++	++++	++	++++
<i>Pinnularia acoricola</i> Hust.			+		
<i>P. borealis</i> Ehr.	+		+	+	+
<i>P. braunii</i> (Grun.) Cl.				+	

Table 1 (contd.)

1	2	3	4	5	6
<i>P. gibba</i> Ehr.		+	+	+	+
<i>P. gibba</i> var. <i>subundulata</i> Mayer				+	
<i>P. globiceps</i> var. <i>krookei</i> Grun.					+
<i>P. interrupta</i> W. Sm.	+	+	+		+
<i>P. lata</i> (Bréb.) W. Sm.		+		+	
<i>P. legumen</i> Ehr.		+			
<i>P. maior</i> (Kütz.) Cl.		+		+	
<i>P. maior</i> var. <i>lacustris</i> Meist.		+			
<i>P. mesolepta</i> (Ehr.) W. Sm.		+		+	
<i>P. mesolepta</i> var. <i>angusta</i> Cl.	+	+			
<i>P. microstauron</i> (Ehr.) Cl.	+	+		+	+
<i>P. microstauron</i> f. <i>biundulata</i> O. Müll.		+			
<i>P. microstauron</i> var. <i>brebissonii</i> (Kütz.) Hust.		+	+	+	+
<i>P. nobilis</i> Ehr.		+	+		
<i>P. platycephala</i> (Ehr.) Cl.		+			
<i>P. viridis</i> (Nitzsch) Ehr.	+	+	+	+	+
<i>P. viridis</i> var. <i>intermedia</i> Cl.	+				
<i>Rhoicosphenia curvata</i> (Kütz.) Grun.		+	+	+	+
<i>Rhopalodia gibba</i> (Ehr.) O. Müll.		+			+
<i>Stauroneis acuta</i> W. Sm.		+	+		+
<i>S. anceps</i> Ehr.	+	+	+		+
<i>S. anceps</i> f. <i>gracilis</i> (Ehr.) Cl.				+	+
<i>S. anceps</i> f. <i>linearis</i> (Ehr.) Cl.	+	+			
<i>S. phoenicentron</i> Ehr.	+	+	+	+	+

Table 1 (contd.)

1	2	3	4	5	6
<i>S. pygmaea</i> Krieg.		++		+	
<i>S. smithii</i> Grun.			+	++	++++
<i>Stephanodiscus astra</i> var. <i>minutulus</i> (Kütz.) Grun.			+		
<i>S. hantzschii</i> Grun.		+	+++	+	+++
<i>Surirella angustata</i> Kütz.	+++	+++	++	++	++
<i>S. biseriata</i> Bréb.		+++	++	+	+
<i>S. biseriata</i> var. <i>bifrons</i> (Ehr.) Hust.			+	+	
<i>S. biseriata</i> var. <i>bifrons</i> f. <i>punctata</i> Meist.					+
<i>S. biseriata</i> var. <i>rostrata</i> Schulz			+		
<i>S. elegans</i> Ehr.		+			
<i>S. linearis</i> W. Sm.	+			+	+
<i>S. linearis</i> var. <i>constricta</i> (Ehr.) Grun.		+	+	+	
<i>S. ovalis</i> Bréb.				+	
<i>S. ovata</i> Kütz.	++++	+++		+++	++++
<i>S. ovata</i> var. <i>pinnata</i> (W. Sm.) Hust.	+	+			
<i>S. ovata</i> var. <i>pseudopinnata</i> Mayer					+
<i>S. robusta</i> var. <i>splendida</i> (Ehr.) VH					+
<i>S. tenera</i> Greg.			+	+	
<i>S. turgida</i> W. Sm.				+	
<i>Synedra acus</i> Kütz.	+	+	++	+	
<i>S. capitata</i> Ehr.		+			
<i>S. parasitica</i> (W. Sm.) Hust.		+	+++		+
<i>S. parasitica</i> var. <i>subcon-</i> <i>stricta</i> Grun.				++	++
<i>S. pulchella</i> (Ralfs) Kütz.		+	++		+
<i>S. rumpens</i> Kütz.			+	+	
<i>S. ulna</i> (Nitzsch.) Ehr.	++++	++++	++++	++++	++++

Table I (contd.)

1	2	3	4	5	6
<i>S. ulna</i> var. <i>aequalis</i> (Kütz.) Hust.	+	+			
<i>S. ulna</i> var. <i>biceps</i> (Kütz.) Schönf.	+	+	+	+	++
<i>S. ulna</i> var. <i>danica</i> (Kütz.) Grun.		+			
<i>S. ulna</i> var. <i>oxyrhynchus</i> (Kütz.) VH	+		+++		++
<i>S. ulna</i> var. <i>spathulifera</i> Grun.	+		+		
<i>Tabellaria fenestrata</i> (Lyngb.) Kütz.					+
<i>T. flocculosa</i> (Roth.) Kütz.		++++			
<i>Xanthophyceae</i>					
<i>Tribonema viride</i> Pasch.		+	+		
<i>Chlorophyta</i>					
<i>Actinastrum hantzschii</i> Lagh.	+		+		
<i>Ankistrodesmus gracilis</i> (Reinsch) Korš.			+		
<i>Chlamydomonas</i> sp.	++	+	+++		+
<i>Cladophora glomerata</i> (L.) Kütz.			++	+	+
<i>Closterium acerosum</i> (Schränk) Ehr.	+++			+	
<i>C. ehrenbergii</i> Menegh.	++		+	+	+
<i>C. leiblenii</i> Kütz.			+++	+	+++
<i>C. lineatum</i> Ehrenb.				+	+
<i>C. littorale</i> Gay					+
<i>C. lunula</i> (O. Müll.) Nitzsch	+				+
<i>C. parvulum</i> Näg.	+			+	
<i>C. rostratum</i> Ehrenb.				+	
<i>C. strigosum</i> Bréb.			++	+	
<i>Coelastrum cambricum</i> Arch.		+			
<i>C. microporum</i> Näg.		++		++	
<i>C. reticulatum</i> (Dang.) Senn.		+			

Table 1 (contd.)

1	2	3	4	5	6
<i>Cosmarium botrytis</i> Menegh.				+	+
<i>C. reniforme</i> Arch.				+	
<i>Crucigenia quadrata</i> Morren		+	+		
<i>C. tetrapedia</i> (Kirchn.) W. et G. S.			+		
<i>Euastrum dubium</i> Näg.		+			
<i>Microspora amoena</i> (Kütz.) Ra- benhorst		+		+	
<i>M. quadrata</i> Hasen	+			+	
<i>M. stagnorum</i> (Kütz.) Lagerh.	+				
<i>Monoraphidium contortum</i> (Thur.) Kom. Legn.				+	
<i>Oedogonium</i> sp.			+	+	
<i>Pandorina morum</i> (O. F. Mül- ler) Bory			+	+	
<i>Pediastrum biradiatum</i> Meyen		+			
<i>P. boryanum</i> (Turp.) Menegh.	+	+	+	+	+
<i>P. duplex</i> Meyen		+	+	+	+
<i>P. tetras</i> (Ehr.) Ralfs		+			
<i>Scenedesmus abundans</i> (Kirchn.) Chod.	+		+	+	
<i>S. acuminatus</i> (Lagh.) Chod.	+		+	+	
<i>S. acutiformis</i> Schröd.			+	+	
<i>S. acutus</i> Meyen			+	+	+
<i>S. brasiliensis</i> Bohl.	+			+	
<i>S. denticulatus</i> Lagerh.			+		
<i>S. disciformis</i> (Chod.) Fott et Kom.			+		
<i>S. intermedius</i> Chod.	+				
<i>S. obliquus</i> (Turp.) Kütz.				+	
<i>S. opoliensis</i> P. Richt.	+				
<i>S. quadricauda</i> (Turp.) Bréb.	+	+	+	+	+
<i>S. velitaris</i> Kom.		+	+		+
<i>Spirogyra</i> sp.	+	+		+	
<i>Staurostrum gracile</i> Ralfs		+			

Table I (contd.)

1	2	3	4	5	6
<i>S. punctulatum</i> Bréb.					+
<i>Stigeoclonium tenue</i> Kütz.	+				
<i>Stigeoclonium</i> sp.	+				
<i>Tetraedron incus</i> (Teil.) G. M. Sm.		+			
<i>Tetrastrum staurogeniaeforme</i> (Schröd.) Lemm.			+		
<i>Ulothrix</i> sp.	+		+		
<i>Vaucheria</i> sp.		+	+	+	+
<i>Rhodophyta</i>					
<i>Chantransia chalybaea</i> (Roth) Fries		+	+	+	+

The smallest number of sessile algae taxa was observed in the Rakówka River (Fig. 1), where especially low number of diatom taxa was found (Tab. II). The lower sections of other investigated tributaries of the Widawka River have similar number of sessile algae. Number of species in these rivers is considerable.

Table II

Number of taxa of the sessile algae in lower sections
of main tributaries of the Widawka River

Group	River				
	Rakówka	Pilsia	Krasawa	Grabia	Nieciecz
<i>Cyanophyta</i>	10	6	16	10	8
<i>Pyrrophyta</i>			2		
<i>Euglenophyta</i>	8	8	7	6	10
<i>Chrysophyceae</i>	1	2			
<i>Bacillariophyceae</i>	113	174	173	179	202
<i>Xanthophyceae</i>		1	1		
<i>Chlorophyta</i>	19	18	23	24	14
<i>Rhodophyta</i>		1		1	
Total	151	210	222	220	234

A similar number of species was observed in the lower section of the Widawka River (L i g o w s k i in press b). No major differences in the number of identified algae according to size of the river were revealed.

Diatoms appear to be a dominant algae in all the rivers studied. They constituted 74.8% of all species in the Rakówka River, 82.5% in the Pilsia River, 77.6% in the Krasawa River, 81.4% in the Grabia River, and 86.3% in the Nieciecz River. These values correspond to those met in other rivers in Poland. Such diatoms as *Navicula hungarica* var. *capitata* and *Synedra ulna* can be found in all samples, and *Cymbella ventricosa*, *Meridion circulare* and *Opephora martyi* in almost all of the samples.

The number of algal taxa in lower sections of some rivers in the Central Poland is presented in Tab. III. In most rivers, the number of taxa of algae ranged from 195 to 263. The smallest number of taxa among various investigated rivers was discovered in the most polluted lower sections of the Rakówka River and the Bzura River. A high number of taxa identified in the Bobrówka River results from a considerable number of collected samples and from the drifting of pond algae to the river.

Table III

Number of taxa of the sessile algae in lower sections
of some rivers in Central Poland

River	Number of taxa
Bobrówka (Ligowski in press a)	373
Bzura (Rakowska 1976)	180
Grabia	220
Krasawa	222
Luciąża (Kalinowska-Kucharska 1984 b)	238 (diatoms only)
Nieciecz	234
Pilsia	210
Rakówka (Kadłubowska et al. 1981)	160
Rakówka	151
Rawka (Rakowska 1984)	263
Strawa (Kalinowska-Kucharska 1984 a)	237 (diatoms only)
Widawka (Ligowski in press b)	195

It is interesting to note permanent presence of *Sphaerotilus natans* in the Rakówka River growing on stones and other objects on the river bottom. In spring also *Zoogloeae ramigera* could be found there. (Tab. I). According to saprobic systems (Zelinka and Marvan 1961 quoted after F o t t 1971, Fjerdingsstad 1964 quoted after E t t l 1980) *Zoogloeae ramigera* is an indicator of the polysaprobic zone, while *Sphaerotilus natans* develops in flowing polysaprobic waters with a small amount of oxygen. The presence of these bacteria is accompanied by algal community with *Euglena* spp. and *Nitzschia palea*. T u r o b o y s k i (1979) believes that a community of diatoms growing with *Sphaerotilus natans* evidences that waters belong to α -mesosaprobic zone. Kolkwitz and Marsson (1908-1910 after P a l u c h 1973) are of the opinion that *Sphaerotilus natans* is an indicator of polysaprobic and α -mesosaprobic zones, and *Zoogloeae ramigera* is an indicator of polysaprobic zone alone. Taking into account the above system an analysis of Tab. I indicates that the lower section of the Rakówka River in May and June should be considered more polluted than in later months. However, all the samples contained a considerable number of photosynthetic plants, which does not allow to classify this water among the most polluted.

The presence of the iron bacterium *Leptothrix ochracea* considered to be a "starving" form of *Sphaerotilus natans* was discovered in the sample taken from the Pilsia River in November. This is a species found in waters poor in organic compounds (P a l u c h 1973). In saprobic systems, this bacterium is considered to be an indicator of the oligosaprobic zone (P a l u c h 1973) similar to red alga *Chantransia chalybaea* (F o t t 1971) found also in this river. It is only in this river that *Tabellaria flocculosa*, another indicator of oligosaprobic waters (F o t t 1971) can be found. The permanent presence of single diatom specimens may be however caused by the drifting of their solid frustules by water currents in the river. *Chantransia chalybaea* was also found in the lower section of the Grabia River (Tab. I) and at various stations of the Widawka River (L i g o w s k i in press b).

Vaucheria sp. growing abundantly on the concrete bottom of the Widawka River (K a d ł u b o w s k a et al. 1981, L i g o w s k i in press b) was found in lower sections of its main

tributaries in the Pilsia River, the Krasawa River, the Grabia River, and the Nieciecz River (Tab. I). *Vaucheria* sp. is considered to be an indicator of β -mesosaprobic zone (Fott 1971) or oligosaprobic zone (Fjerdinstad after Ettl 1980). The absence of this taxon in the lower section of the polluted Rakówka River is concordant with these opinions.

Several rare species of algae were found in the Widawka River basin. They will be analyzed in a separate study.

4.2. Quantitative composition

The largest number of cells was found in the lower section of the Rakówka River - on the average 8.1×10^6 per 1 cm^3 . The smallest number of cells was observed in November - 0.5×10^6 per 1 cm^3 , and the biggest in May - 23×10^6 per 1 cm^3 (Fig. 2). Together with algae, the bottom was covered by very abundant bacteria *Sphaerotilus natans* and *Zooglaea ramigera*, which were not counted. It was observed that the number of attached algal cells tended to decline during the vegetation season. Diatoms, among which *Nitzschia palea* constantly dominated, were found in the largest quantities. Blue-green algae belonging to *Oscillatoria* and euglenoid *Euglena* were found to be quite abundant.

In the Pilsia River the smallest number of algal cells was observed. This number ranged here from 0.19×10^6 per 1 cm^3 in July to 0.88×10^6 per 1 cm^3 in June (Fig. 2). In lower section of this river, the average number of 0.6×10^6 attached algae cells per 1 cm^3 was found. Among the dominant species were diatoms of the genus *Fragilaria* and mainly - *Fragilaria construens* with its varieties.

The average number of 6.7×10^6 cells per 1 cm^3 was observed in the lower section of the Krasawa River. The smallest number of algae cells per 1 cm^3 was discovered in July - 4.0×10^6 and the largest in June - 9.2×10^6 (Fig. 2). In late spring (May and June), apart from the dominant diatoms, there could be found blue-green algae from the genus *Oscillatoria* in large quantities. Among diatoms, *Navicula cryptocephala* and species from the genus *Fragilaria* were dominant.

In the Grabia River, the smallest number of cells was ob-

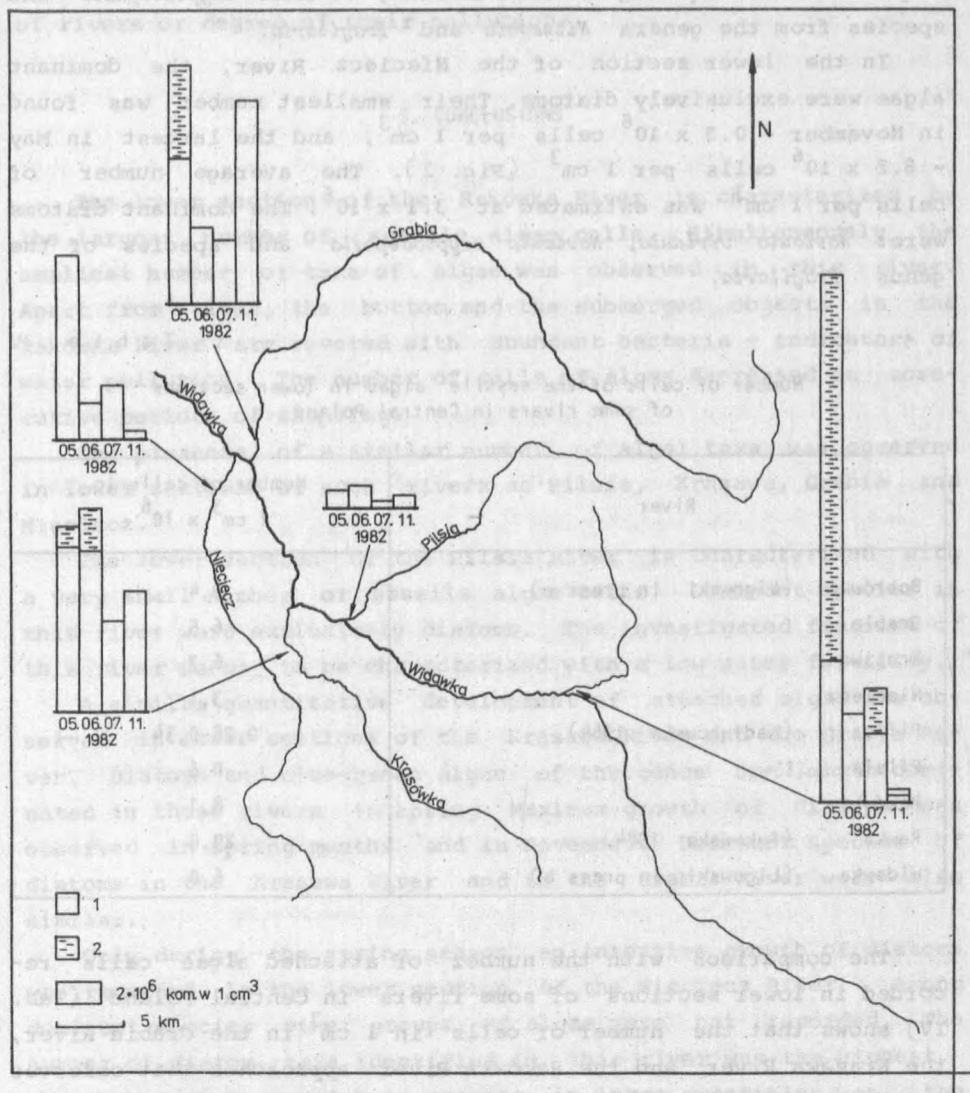


Fig. 2. Number of cells of the sessile algae in lower sections of the main tributaries of the Widawka River

served in July - 2.8×10^6 per 1 cm³, and the largest in May - 10.7×10^6 per 1 cm³ (Fig. 2). On the average the number of cells amounted to 6.5×10^6 per 1 cm³. Blue-green alga *Oscillatoria limosa* was dominant in May (40.5% of the total number of cells) and in the remaining months - diatoms, mainly: *Navicula*

hungarica var. *capitata*, *Navicula viridula*, *Navicula cryptocephala* and species from the genera *Nitzschia* and *Fragilaria*.

In the lower section of the Nieciecz River, the dominant algae were exclusively diatoms. Their smallest number was found in November - 0.3×10^6 cells per 1 cm^3 , and the largest in May - 8.2×10^6 cells per 1 cm^3 (Fig. 2). The average number of cells per 1 cm^3 was estimated at 3.1×10^6 . The dominant diatoms were: *Navicula viridula*, *Navicula cryptocephala* and species of the genus *Fragilaria*.

Table IV

Number of cells of the sessile algae in lower sections
of some rivers in Central Poland

River	Number of cells in $1 \text{ cm}^3 \times 10^6$
Bobrówka (Ligowski in press a)	9,4
Grabia	6,5
Krasawa	6,7
Nieciecz	3,1
Pilica (Kadłubowska 1964)	0,28-0,34
Pilsia	0,6
Rakówka	8,1
Rawka (Rakowska 1984)	29,0
Widawka (Ligowski in press b)	6,0

The comparison with the number of attached algae cells recorded in lower sections of some rivers in Central Poland (Tab. IV) shows that the number of cells in 1 cm^3 in the Grabia River, the Krasawa River and the Rakówka River approaches that observed in the Bobrówka River (Ligowski in press a) and the Widawka River (Ligowski in press b). An exceptionally small number of cells per 1 cm^3 was observed in the Pilica River (Kadłubowska 1964) and the Pilsia River. The largest number of cells per 1 cm^3 is reported by Rakowska (1984) from the Rawka River - a right side tributary of the Bzura River. This comparison seems to indicate that the number of algal

cells in the rivers under comparison does not depend on the size of rivers or degree of their pollution.

5. CONCLUSIONS

The lower section of the Rakówka River is characterized by the largest number of sessile algae cells. Simultaneously the smallest number of taxa of algae was observed in this river. Apart from algae, the bottom and the submerged objects in the Rakówka River are covered with abundant bacteria - indicators of water pollution. The number of cells of algae decreased in consecutive periods of sampling.

The presence of a similar number of algal taxa was observed in lower sections of such rivers as Pilsia, Krasawa, Grabia and Nieciecz.

The lower section of the Pilsia River is characterized with a very small number of sessile algae cells. Dominant species in this river were exclusively diatoms. The investigated fragment of this river seems to be characterized with a low water fertility.

A similar quantitative development of attached algae was observed in lower sections of the Krasawa River and the Grabia River. Diatoms and blue-green algae of the genus *Oscillatoria* dominated in these rivers in spring. Maximum growth of diatoms was observed in spring months and in November. Dominant species of diatoms in the Krasawa River and in the Grabia River were also similar.

Only during the spring season an intensive growth of diatoms was recorded in the lower section of the Nieciecz River. Among dominant species other groups of algae were not recorded. The number of diatom taxa identified in this river was the highest.

Vaucheria sp., which is present in large quantities on the concrete bottom of the Widawka River (K a d ł u b o w s k a et al. 1981, L i g o w s k i in press b) was observed in lower sections of such rivers as Pilsia, Krasawa, Nieciecz, and Grabia in small quantities on the solid substrate like stones and piles. The presence of *Vaucheria* sp. was not discovered in the most polluted river of the Widawka basin - Rakówka.

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7. STRESZCZENIE

Badania glonów osiadłych głównych dopływów Widawki podjęte zostały w związku ze zmianą warunków hydrologicznych na tym obszarze, spowodowaną przez odkrywkę węgla brunatnego Bełchatów. Na podstawie prób zebranych w maju, czerwcu, lipcu i listopadzie 1982 r. z ujściowych odcinków Rakówki, Pilsa, Krasawy, Grabi i Niecieczy (rys. 1) określono skład jakościowy i ilościowy glonów osiadłych. Zidentyfikowano 417 taksonów glonów, z czego 313 należało do okrzemek oraz 3 gatunki bakterii (tab. I). Najczęściej występującymi glonami są okrzemki: *Navicula hungarica* var. *capitata*, *Synedra ulna*, *Cymbella ventricosa*, *Meridion circulare* i *Opephora martyi*. Najmniejszą liczbę gatunków stwierdzono w ujściowym odcinku Rakówki (tab. II). Jednocześnie w tej rzece występowała największa liczba komórek glonów (rys. 2). Fakty te oraz występowanie indykatorów wód α -mezosaprobnych i polisaprobnych wskazują na zanieczyszczenie ujściowego odcinka Rakówki. W składzie ilościowym we wszystkich rzekach dominowały okrzemki (rys. 2). W Krasawie, Grabi i Niecieczy obserwowano wiosenne maksimum rozwojowe glonów. Pilsia charakteryzuje się małą liczbą komórek glonów osiadłych w jej odcinku ujściowym.

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